

Beneficial Properties of Insects:

Antimicrobial Peptides, Chitin, and Lauric Acid

Insect based ingredients are becoming widely accepted and utilized in both livestock feeds and pet foods. Insects are highly nutritious, palatable and sustainable. They are intentionally and opportunistically ingested by cats, dogs, poultry, swine, fish and other species including grazing livestock. The three most commonly used insect species as ingredients in animal feed and pet food are crickets (*Acheta domesticus*), larvae of the black soldier fly (*Hermetia illucens*, BSFL), and mealworms (*Tenebrio molitor*). As highlighted by Koutsos et al. (2022), there are numerous studies demonstrating the nutritional efficacy of insect ingredients, many of which have focused on BSFL. Recently, researchers have begun to explore specific properties of BSFL that may provide animal health benefits in addition to nutritional benefits. These properties include antimicrobial peptides (AMPs), chitin, and lauric acid.

Antimicrobial Peptides (AMPs)

Insects display a particularly strong resistance to most infections. One reason for this resistance is the unique insect immune system that is dominated by robust innate immune defenses including

antimicrobial peptides (Samakovlis et al., 1990). These peptides may also be referred to as host defense peptides, and are present in many different organisms and ecosystems (Kim et al., 2005). AMPs act by preventing harmful pathogens from infecting the host (Harlystiarini et al., 2019). This efficacy exists against a range of pathogen types including viruses, bacteria, fungi, and parasites (Bahar et al., 2013). There are many different mechanisms by which AMPs may impact microorganisms; however, most AMPs act directly on the pathogen by disrupting the cellular membrane or preventing the cell from replicating. This direct interaction method of antimicrobial activity makes it incredibly difficult for pathogens to develop resistance to AMPs, and in 2014, Bagley et al. proposed the use of AMPs as an alternative to antibiotics in an effort to combat concerns regarding antibiotic resistance.

AMPs have demonstrated a variety of properties that could be beneficial to the animals. Wang et al. (2022) used meta-analysis to demonstrate the ability of AMPs to boost aquatic species health when fed as part of the diet. This was shown in both

fresh and saltwater aquatic species. Additionally, AMPs extracted from the gastrointestinal tract of swine have been shown to alleviate growth depression in poultry exposed to heat stress, and increase the populations of beneficial bacteria such as *Lactobacillus* spp in the gut (Hu et al., 2017; Daneshmand et al., 2019). AMPs are also very small molecules, making them more resistant to high temperature protein degradation and conformational changes, making them an excellent candidate for animal feed applications, which are often exposed to high temperatures during extrusion or pelleting (Qiao et al., 2020).

Being an integral part of their robust and unique immune system, AMPs have been identified in a variety of insect species. There are 57 identified genes coding for various AMPs present in BSFL adults and larvae (Vogel et al., 2018; Moretta et al., 2020). This is the highest reported number of AMP genes of any invertebrate (Vogel et al., 2018). Although the abundance of these AMP coding genes in BSFL is clear, the expression of these genes and type of AMPs produced can be modified. BSFL rearing practices such as diet, processing method, and breeding program will impact AMP expression and efficacy against pathogens (Kong et al., 2019). This high level of AMP

expression makes BSFL ingredients a beneficial addition to any animal feeding program.

Chitin

Chitin is a highly abundant compound in nature that is a component of insect and crustacean exoskeletons (Kumar et al., 2000). Chitin was previously thought of as an anti-nutritional component of animal diets due to its ability to decrease digestibility when fed at high concentrations. However, as chitin research has developed, the compound is now known to improve disease response by priming the immune system when fed at less than 3% of total diet composition (Elieh et al., 2018). The mechanism behind this priming activity is thought to be multifaceted. First, chitin has the ability to induce a number of adaptive immune pathways in both human and animal species including Th1, Th2, and Th17 pathways (Da Silva et al., 2010). These pathways are critical mediators of gut inflammation and cellular signaling. Additionally, in many species, chitin is not absorbed in the small intestine, allowing it to pass to the large intestine where bacteria can utilize the compound as a fermentation substance (Yu et al., 2019).

Chitin has also proven effective as a prebiotic, promoting increased proliferation

of beneficial lactic acid bacteria, and a decreased occurrence of pathogenic bacteria in the gut such as *E. coli* and *Salmonella* spp. (Dörper et al., 2021). Chitin derived from insect species has been shown to improve performance parameters and carcass quality in poultry and reduce post weaning diarrhea in piglets when included in the diet at low levels (Lokman et al., 2019; Xu et al., 2018). Shah et al (2022) concluded that chitin has the potential to improve growth and milk production in dairy cows, and even increased wool production in small ruminants. Feeding chitin at low inclusion rates also enhanced butyrate production in the gut. This volatile fatty acid is speculated to play a critical role in developing healthy microflora in the animal gastrointestinal tract (Khempaka et al., 2011).

Lauric Acid

The third beneficial component of BSFL derived ingredients is lauric acid. Lauric acid is a medium chain fatty acid (MCFA) commonly sourced from coconut and palm oils. Lauric acid is the major component of BSFL lipid fraction, and has shown a wide array of antimicrobial properties (Kumar et al., 2020). It is important to keep in mind that the lauric acid content of BSFL is somewhat diet dependent, with

concentrations varying based on the feedstock used in BSFL rearing.

Similar to AMPs, lauric acid and its derivatives have been proposed by Borrelli et al. (2021) as an alternative to antibiotics in order to combat antimicrobial resistance. Generally speaking, lauric acid disrupts the cellular membrane by inducing cell lysis. This is a generalized response, which makes it difficult for bacteria and other pathogens to develop resistance (Jackman et al., 2020). Lauric acid is highly bioavailable, being easily absorbed and transported throughout the body unlike many other fatty acids (Dayrit et al., 2015). This increased viability is because lauric acid does not require any lipolysis before being absorbed by intestinal cells (Greenberger et al., 1965; Guillot et al., 1993). Because of the ease at which lauric acid can be absorbed, Hall et al. (2014) shown as an excellent feed component for geriatric animals, reducing the effects of age on kidney function and other biomarkers .

In animal studies, BSFL oil has demonstrated improved growth performance in newly weaned piglets when compared to a diet containing poultry fat (Van Heugten et al., 2022). In rainbow trout, BSFL oil decreased gut

inflammation and improved innate immune response when compared to a soybean-based control diet (Kumar et al., 2021). In poultry, the inclusion of BSFL oil reduced levels of harmful bacteria and decreased the expression of inflammatory markers in the digestive tract (Sypniewski et al., 2020).

Conclusions

Insect-derived ingredients offer a wide range of value-added properties that can benefit animal nutrition and performance. Of the many beneficial compounds found in insect ingredients, AMPs, chitin, and lauric acid are the best characterized and most understood. All three of these compounds have displayed antimicrobial potential, and are at relatively high concentrations in BSFL-derived ingredients when compared to other sources.

AMPs, chitin, and lauric acid have shown beneficial properties when fed to a multitude of animal species. The combination of these three beneficial compounds in BSFL ingredients make it an excellent ingredient choice for animal feeding programs through decreased mortality, inflammation reduction, improved gut microflora, and improved growth performance.

References

- Barragan-Fonseca, K. B., M. Dicke, and J. J. A. van Loon. 2017. Nutritional value of the black soldier fly (*Hermetia illucens* L.) and its suitability as animal feed - A review. *J. Insects Food Feed*, 3(2):105-120. doi:10.3920/JIFF2016.0055
- Bagley, C. P. (2014). Potential role of synthetic antimicrobial peptides in animal health to combat growing concerns of antibiotic resistance-a review. *Wyno Academic Journal of Agricultural Sciences*, 2(2), 19-28.
- Bahar, A. A., & Ren, D. (2013). Antimicrobial peptides. *Pharmaceuticals*, 6(12), 1543-1575.
- Beutler, B. (2004). Innate immunity: an overview. *Molecular immunology*, 40(12), 845-859.
- Bonilla, F. A., & Oettgen, H. C. (2010). Adaptive immunity. *Journal of Allergy and Clinical Immunology*, 125(2), S33-S40.
- Borrelli, L., Varriale, L., Dipineto, L., Pace, A., Menna, L. F., & Fioretti, A. (2021). Insect derived lauric acid as promising alternative strategy to antibiotics in the antimicrobial resistance scenario. *Frontiers in Microbiology*, 12, 620798.
- Daneshmand, A., Kermanshahi, H., Sekhavati, M. H., Javadmanesh, A., & Ahmadian, M. (2019). Antimicrobial peptide, cLF36, affects performance and intestinal morphology, microflora, junctional proteins, and immune cells in broilers challenged with *E. coli*. *Scientific reports*, 9(1), 1-9.
- Da Silva, C. A., Pochard, P., Lee, C. G., & Elias, J. A. (2010). Chitin particles are multifaceted immune adjuvants. *American journal of respiratory and critical care medicine*, 182(12), 1482-1491.
- Dayrit, F. M. (2015). The properties of lauric acid and their significance in coconut oil. *Journal of the American Oil Chemists' Society*, 92(1), 1-15.
- Dörper, A., Veldkamp, T., & Dicke, M. (2021). Use of black soldier fly and house fly in feed to promote sustainable poultry production. *Journal of Insects as Food and Feed*, 7(5), 761-780.
- Elieh Ali Komi, D., Sharma, L., & Dela Cruz, C. S. (2018). Chitin and its effects on inflammatory and immune responses. *Clinical reviews in allergy & immunology*, 54(2), 213-223.
- Greenberger, N. J., Franks, J. J., & Isselbacher, K. J. (1965). Metabolism of 1-C¹⁴ Octanoic and 1-C¹⁴ Palmitic Acid by Rat Intestinal Slices. *Proceedings of the Society for Experimental Biology and Medicine*, 120(2), 468-472.
- Guillot, E., Vaugelade, P., Lemarchali, P., & Rat, A. R. (1993). Intestinal absorption and liver uptake of medium-chain fatty acids in non-anaesthetized pigs. *British journal of nutrition*, 69(2), 431-442.
- Hall, J. A., Yerramilli, M., Obare, E., Yu, S., & Jewell, D. E. (2014). Comparison of serum concentrations of symmetric dimethylarginine and creatinine as kidney function biomarkers in healthy geriatric cats fed reduced protein foods enriched with fish oil, L-carnitine, and medium-chain triglycerides. *The Veterinary Journal*, 202(3), 588-596.
- Harlystiarini, H., Mutia, R., Wibawan, I. W. T., & Astuti, D. A. (2019). In vitro antibacterial activity of black soldier fly (*Hermetia illucens*) larva extracts against gram-negative bacteria. *Buletin Peternakan*, 43(2), 125-129.
- Hu, F., Gao, X., She, R., Chen, J., Mao, J., Xiao, P., & Shi, R. (2017). Effects of antimicrobial peptides on growth performance and small intestinal function in broilers under chronic heat stress. *Poultry science*, 96(4), 798-806.
- Jackman, J. A., Boyd, R. D., & Elrod, C. C. (2020). Medium-chain fatty acids and monoglycerides as feed additives for pig production: towards gut health improvement and feed pathogen mitigation. *Journal of animal science and biotechnology*, 11(1), 1-15.
- Khempaka, S., Chitsatchapong, C., & Molee, W. (2011). Effect of chitin and protein constituents in shrimp head meal on growth performance, nutrient digestibility, intestinal microbial populations, volatile fatty acids, and ammonia production in broilers. *Journal of Applied Poultry Research*, 20(1), 1-11.
- Kim, J. Y., Park, S. C., Kim, M. H., Lim, H. T., Park, Y., & Hahm, K. S. (2005). Antimicrobial activity studies on a trypsin-chymotrypsin protease inhibitor obtained from potato. *Biochemical and biophysical research communications*, 330(3), 921-927.
- Kong, H., Dong, C., Jing, W., Zheng, M., Tian, Z., Hou, Q., ... & Zhu, S. (2019). Transcriptomic insight into antimicrobial peptide factors involved in the prophylactic immunity of crowded *Mythimna separata* larvae. *Developmental & Comparative Immunology*, 98, 34-41.
- Kumar, M. N. R. (2000). A review of chitin and chitosan applications. *Reactive and functional polymers*, 46(1), 1-27.
- Kumar, V., Fawole, F. J., Romano, N., Hossain, M. S., Labh, S. N., Overturf, K., & Small, B. C. (2021). Insect (black soldier fly, *Hermetia illucens*) meal supplementation prevents the soybean meal-induced intestinal enteritis in rainbow trout and health benefits of using insect oil. *Fish & Shellfish Immunology*, 109, 116-124.
- Lokman, I. H., Ibitoye, E. B., Hezmee, M. N. M., Goh, Y. M., Zuki, A. B. Z., & Jimoh, A. A. (2019). Effects of chitin and chitosan from cricket and shrimp on growth and carcass performance of broiler chickens. *Tropical animal health and production*, 51(8), 2219-2225.
- Qiao, Z., Fu, Y., Lei, C., & Li, Y. (2020). Advances in antimicrobial peptides-based biosensing methods for detection of foodborne pathogens: A review. *Food Control*, 112, 107116.
- Samakovlis, C., Kimbrell, D. A., Kylsten, P., Engström, Å., & Hultmark, D. (1990). The immune response in *Drosophila*: pattern of cecropin expression and biological activity. *The EMBO journal*, 9(9), 2969-2976.
- Shah, A. M., Qazi, I. H., Matra, M., & Wanapat, M. (2022). Role of Chitin and Chitosan in Ruminant Diets and Their Impact on Digestibility, Microbiota and Performance of Ruminants. *Fermentation*, 8(10), 549.
- Van Heugten, E., Martinez, G., McComb, A., & Koutsos, L. (2022). Improvements in Performance of Nursery Pigs Provided with Supplemental Oil Derived from Black Soldier Fly (*Hermetia illucens*) Larvae. *Animals*, 12(23), 3251.
- Vogel, H., Müller, A., Heckel, D. G., Gutzzeit, H., & Vilcinskis, A. (2018). Nutritional immunology: diversification and diet-dependent expression of antimicrobial peptides in the black soldier fly *Hermetia illucens*. *Developmental & Comparative Immunology*, 78, 141-148.
- Wang, J., Wilson, A. E., Su, B., & Dunham, R. A. (2022). Functionality of dietary antimicrobial peptides in aquatic animal health: Multiple meta-analyses. *Animal Nutrition*.
- Xu, Y., Wang, Z., Wang, Y., Yan, S., & Shi, B. (2018). Effects of chitosan as growth promoter on diarrhea, nutrient apparent digestibility, fecal microbiota and immune response in weaned piglets. *Journal of applied animal research*, 46(1), 1437-1442.
- Yu, M., Li, Z., Chen, W., Rong, T., Wang, G., & Ma, X. (2019). *Hermetia illucens* larvae as a potential dietary protein source altered the microbiota and modulated mucosal immune status in the colon of finishing pigs. *Journal of animal science and biotechnolomgeesner@abstractdisplays.comgy*, 10(1), 1-16.